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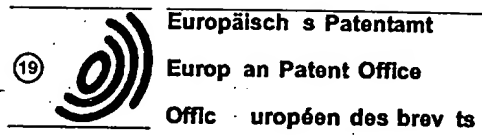
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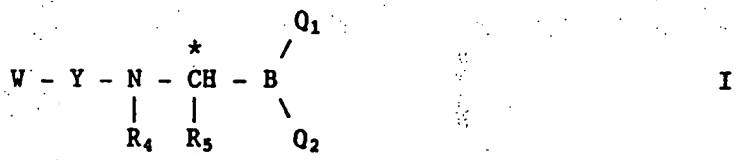
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(54) **Peptide boronic acid derivatives having protease inhibiting activity.**

(57) Borolysine peptidomimetics of formula I



wherein W, Y, R₄, R₅, Q₁ and Q₂ as defined in claim 1 are potent thrombin inhibitors.

EP 0 471 651 A2

This invention relates to inhibitors of serine proteases such as thrombin, factor Xa, kallikrein and plasmin as well as other serine proteases like prolyl endopeptidases and Ig A1 Protease. Thrombin, the last enzyme in the coagulation system, cleaves soluble fibrinogen to fibrin, which is then crosslinked and forms an insoluble gel forming the matrix of a thrombus. When a vessel is damaged, the above process is necessary to stop bleeding. Under normal circumstances there is no measurable amount of thrombin present in plasma. Increase of the thrombin concentration can result in formation of clots, which can lead to thromboembolic disease, one of the most common serious medical problems of our time.

Thrombin contributes to haemostatic control by means of several biological reactions. In addition to its primary function, the conversion of fibrinogen to fibrin, thrombin activates Factor XIII, which is responsible for the crosslinking of fibrin. Thrombin also acts by means of a positive feedback mechanism involving the activation of Factors V and VIII, which both are necessary for its own formation from prothrombin. Thrombin has another essential role: its binding to platelets initiates platelet release and aggregation which is responsible for primary haemostasis.

The reactions of thrombin are further controlled by natural inhibitors in plasma. The most important of these are antithrombin III and heparin. These two compounds have been isolated and are therapeutically and prophylactically used in conditions where there is an imbalance in the haemostatic mechanism with risk for prothrombin activation.

Synthetic thrombin inhibitors, having oral activity, would be useful as alternatives to the parenteral administration of these natural inhibitors. Much research in this area has resulted in the synthesis of good inhibitors of thrombin *in vitro*, but as yet there is no really good candidate for oral therapeutic use. By imitating amino acid sequences of fibrinogen, the important natural substrate of thrombin, several good short peptide substrates for thrombin have been produced. These peptide substrates have also been transformed into inhibitors of the enzyme. Thus, the chromogenic substrates D-Phe-Pro-Arg-pNA and D-Phe-Pip-Arg-PNA mimic the sequence preceding the bond split by thrombin. The corresponding reversible and irreversible inhibitors, D-Phe-Pro-Arginal and D-Phe-Pro-Arg-CH₂Cl show inhibition *in vitro* in the 10⁻⁸ M range.

Chloromethylketones are generally too nonspecifically re-active to be ideal for therapeutic use, and the peptide aldehyde exemplified above has quite a low LD₅₀ value.

Factor Xa is the coagulation enzyme responsible for the generation of thrombin by limited proteolysis of its zymogen, prothrombin. On a weight for weight basis factor Xa is at least 10 times more thrombogenic *in vivo* than thrombin. This arises from the fact that factor Xa is one step above thrombin in the amplifying cascade system, so that one molecule of factor Xa can generate many molecules of thrombin from its precursor. Its propensity may also arise from the rather slow removal of factor Xa by the body. Thrombin, unlike factor Xa, is rapidly cleared from circulating blood onto high affinity sites on the vessel wall. The central position of factor Xa at the junction of the intrinsic and the extrinsic pathways should make it a suitable target for modulating the haemostatic mechanism.

Kallikrein is formed from prekallikrein by the action of factor XII, when located on a negatively charged surface. Kallikrein in turn can cleave factor XII to factor XIIa, thereby forming a reciprocal activation system. Factor XIIa is the first enzyme of the intrinsic part of the coagulation system. The significance of the contact system is probably as a surface mediated defense mechanism, and a large scale activation of the system is normally seen during shock or disseminated intravascular coagulation (DIC). The role of kallikrein at this stage is to cleave high molecular weight kininogen with the release of the vasodilator, bradykinin. Bradykinin also causes increased vascular permeability, pain and migration of the neutrophil leucocytes. Inhibitors of kinin formation have been shown to be beneficial in certain types of inflammation, including arthritis and pancreatitis, and may be useful also in the treatment of asthma. The only substance that has attained clinical significance as a kallikrein inhibitor, is aprotinin (Trasylol). Aprotinin is a polypeptide of molecular weight 6,500, and forms a stable complex with proteases, having a binding constant of 10⁻¹²-10⁻¹³ M.

Fibrinolysis is the process which causes an enzymatic dissolution of fibrinogen and fibrin clots. Plasma contains a protein, plasminogen, which under the influence of various activators is converted to plasmin, a proteolytic enzyme, the activity of which resembles that of trypsin. Plasmin breaks down fibrinogen and fibrin to fibrin/fibrinogen degradation products.

Under normal conditions, the fibrinolysis system is in balance with the coagulation system. Small thrombi formed in the blood stream can be dissolved enzymatically and the circulation through the vessels can be restored by the activation of the fibrinolytic system in the body. If the fibrinolytic activity is too high, it may cause or prolong bleeding. The activity can be inhibited by natural inhibitors in the blood.

Prolyl endopeptidase cleaves peptide bonds on the carboxyl side of proline residues within a peptide chain. It is a serine protease which readily degrades a number of neuropeptides including substance P, neurotensin, thyrotropin-releasing hormone and luteinizing hormone-releasing hormone and which has been associated with the ability of cells to produce interleukin 2 (IL-2). The enzyme is inhibited by benzoyloxycarbonyl-prolyl-proline

with a K_i of 14 nM. Despite the fact that almost nothing is known about the physiological role of prolyl endopeptidases, it may play a prominent role in the regulation of the biological activities of various neuropeptides.

The IgA protease-catalyzed cleavage of IgA, the predominant form of antibody which comprises the first line of defense against infection, separates the Fc from the antigen-binding Fab fragments of the molecule. Such cleavage would be expected to impair or abolish its antimicrobial activity. All IgA proteinases identified thus far cleave after a proline residue within the hinge region of human IgA. Peptide prolyl-boronic acids inhibit IgA 1 proteinases from *Neisseria gonorrhoea* and *Haemophilus influenzae* indicating these enzymes belong to the serine protease family of proteolytic enzymes.

The multiple roles played by thrombin in a variety of physiological processes which have been associated with pathological disorders such as cancer, inflammation and neuronal activity, suggest a potential use of thrombin inhibitors in several indications not strictly related to the cardiovascular system.

Many tumor cells have been shown to elicit procoagulant activity associated with the generation of thrombin. As a consequence, local fibrin deposition and coagulation occur which are thought to be important for the growth of the tumor. Additionally, due to its effects on endothelial cells, thrombin may facilitate the extravasation of tumor cells during metastasis. Hence, thrombin inhibitors may prove beneficial not only in the treatment of certain cancers but also in reducing the hypercoagulability frequently observed in patients during therapy with chemotherapeutic agents.

Thrombin activation of endothelial cells induces a number of pro-inflammatory changes such as synthesis and release of interleukin-1, prostaglandins and platelet-activating factor. Additionally, thrombin induces the exposure of GMP-140 and CD63, two adhesive molecules responsible for the adhesion of leukocytes to the endothelial surface. Thrombin also increases the vascular permeability to proteins, an action which involves neutrophils, and cleaves the interleukin-8-precursor protein, a peptide supposed to be involved in respiratory disorders, rheumatoid arthritis and ulcerative colitis.

Its involvement in all these pro-inflammatory processes makes thrombin to a potential target for the therapeutic treatment with thrombin inhibitors of inflammation-related pathological disorders.

The activity of the protease nexin-1, a modulator of nerve growth and a specific natural thrombin antagonist, is markedly and specifically decreased in patients with Alzheimer's disease. This, together with the observation that thrombin-like activity was increased in the Alzheimer's brains, suggests that thrombin inhibitors may have potential for limiting or reversing neuronal pathological changes associated with thrombin hyperactivity.

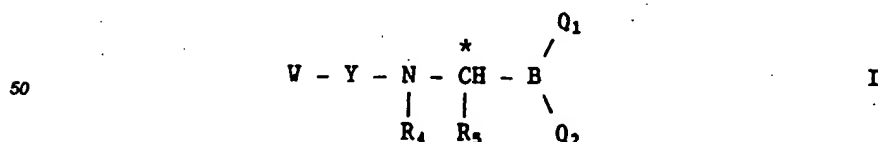
Boronic acids have been studied as inhibitors of various serine esterases and proteases. The first boronic acid-containing amino acid analog to be used as a protease inhibitor was the boronic acid analog of N-acetyl-L-phenylalanine, which was used as an inhibitor of chymotrypsin and subtilisin. Peptide boronic acids have been used as inhibitors of chymotrypsin, subtilisin, and elastases.

The interaction of boronic acids with proteases in biological systems is known and various simple boronic acids have been shown to be sufficiently nontoxic for use in humans. Peptide boronic acid inhibitors of elastase have recently been used in animal trials in relation to emphysema. Unlike the peptide chloromethylketones, there was no toxicity reported at biologically effective dosage levels.

European Patent Application 293 881 describes the preparation of peptides comprising C-terminal boronic acid derivatives of lysine, ornithine and arginine, and their use as inhibitors of trypsin-like serine proteases. The other amino acids in the peptides are all either the D- or the L- forms of the 20 naturally-occurring amino acids.

It has now been found that compounds, having superior properties as inhibitors of trypsin-like serine proteases, are obtained when the peptide contains at least one unnatural α -amino acid having a hydrophobic side chain.

Accordingly, the present invention provides compounds of formula I



wherein W = H or an N-protecting group;

Y is a sequence of n amino acids selected such that the n+1 amino acid peptide Y-Lys or Y-Arg has an affinity for the active site of a trypsin-like protease, where n is an integer from 1 to 10 preferably 1 to 4, and in which

at least n amino acid is an unnatural amino acid having a hydrophobic side chain;
 Q₁ and Q₂, which may be the same or different, are selected from -OH, -COR₁, -CONR₁R₂, -NR₁R₂, and -OR₃,
 or Q₁ and Q₂ taken together form a diol residue; R₁, R₂ and R₃, which may be the same or different, are C₁₋₁₀
 alkyl, C₆₋₁₀ aryl, C₆₋₁₀ aralkyl, or phenyl substituted by up to three groups selected from C₁₋₄ alkyl, halogen and
 C₁₋₄ alkoxy;

R₄ is hydrogen or C₁₋₁₀alkyl R₅ is a group -A-X wherein A is

-(CH₂)_z- in which z is 2,3,4 or 5; -CH(CH₃)-(CH₂)₂-;
 -CH₂-CH(CH₃)-CH₂-; -(CH₂)₂-CH(CH₃)-; -(CH₂)₂-C(CH₃)₂-;
 -CH(CH₃)-(CH₂)₃-; -CH₂-CH(CH₃)-(CH₂)₂-; -CH₂-CH₂-CH(CH₃)-CH₂;
 (CH₂)₃-CH(CH₃)-; -(CH₂)₃-C(CH₃)₂-; C₆₋₁₀aryl, C₆₋₁₀aralkyl

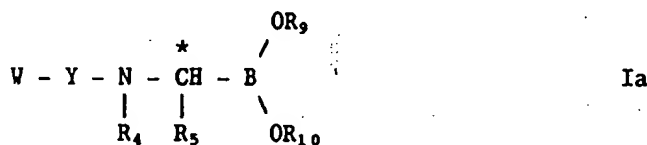
and X is -NH₂, -NH-C(NH)-NH₂, -S-C(NH)-NH₂, -N₃, C₁₋₄ alkoxy, C₁₋₄ alkylthio, or -Si(CH₃)₃ or R₄ and R₅
 together form a trimethylene group and the asymmetric carbon atom marked * may have the D- or L-configuration,
 or represent any mixture of these.

By an unnatural amino acid is meant any amino acid other than D- or L- Ala, Arg, Asp, Cys, Gln, Glu, Gly,
 His, Ile, Leu, Lys, Met, Phe, Pro, Ser, Thr, Trp, Tyr, or Val.

Preferably the N-protecting group W is of formula H(CH₂CH₂O)_p- where p = 3-30; R₆CO-; R₇OCO- or R₈SO₂-
 in which R₆ is C₁₋₆ alkyl; R₇ is C₁₋₆ alkyl, phenyl, benzyl or naphthyl; and R₈ is phenyl, naphthyl or C₁₋₄ alkyl-
 phenyl; of which R₇OCO- is preferred. The most preferred protecting groups are those of formula R₇'OCO- in
 which R₇' is tert-butyl (designated Boc), and in which R₇' is benzyl (designated Z).

Preferably R₆ is R₅' where R₅' is -(CH₂)_z-X', in which X' is -NH₂, -NH-C(NH)-NH₂, -N₃ or -Si(CH₃)₃, and z'
 is 2,3 or 4.

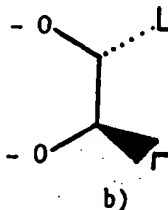
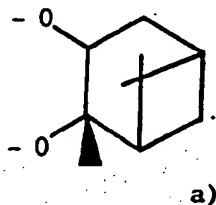
Preferred compounds are those of formula Ia



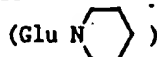
wherein: W, Y, R₄ and R₅ are as defined above and R₉ and R₁₀ represent the residue of a dihydroxy compound.

Useful examples are, 2,3-butanediol; catechol; 2,3-dimethylbutandiol-2,3; cyclohexanediol; ethylene
 glycol; 1,2-hexanediol; 2,3-hexanediol; diethanolamine or aliphatic or aromatic compounds having hydroxy
 groups that are substituted on adjacent carbon atoms or on carbon atoms substituted by another carbon atoms.

Particularly preferred are those compounds in which Q₁ and Q₂ taken together represent the group OPin
 of formula a) or the group of formula b)



The amino acids constituting Y are α-amino acids which may be selected from the L-amino acids naturally
 occurring in proteins, their corresponding enantiomeric D-amino acids or chemically modified alpha-amino
 acids such as glutamic acid gamma-piperidide

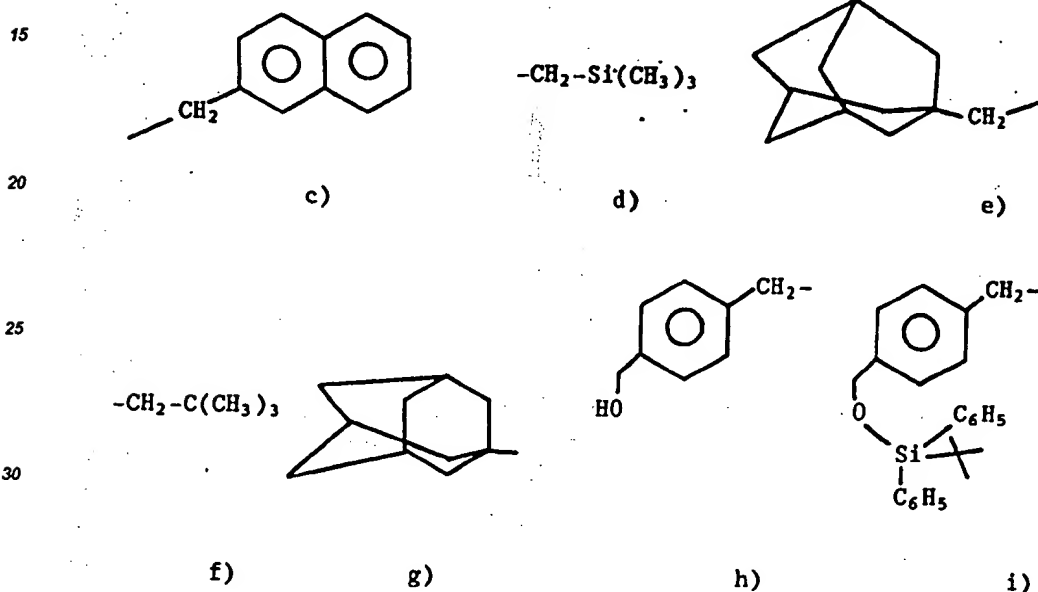


or pIpecolic acid (Pip), provided that at least one amino acid is an unnatural amino acid having a hydrophobic
 side chain.

Preferred unnatural amino acids are of formula II

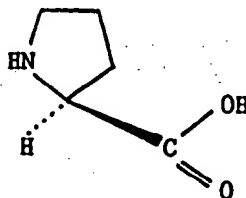


in which R₁₁ is a hydrophobic group. Preferred hydrophobic groups consist of a methylene group linked to an optionally by a polar group substituted aromatic group or an alicyclic group having at least two rings and no polar substituents, or to a tert. butyl or trimethylsilyl group. Preferably R₁₁ is R₁₁' where R₁₁' is a group of formulae c), d), e), f), g), h) or l)

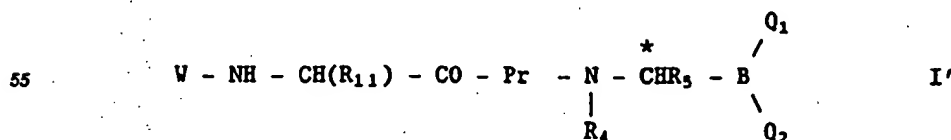


The unnatural amino acids of formula II may be in D- or L- form or any mixture of these, but are preferably in D-form.

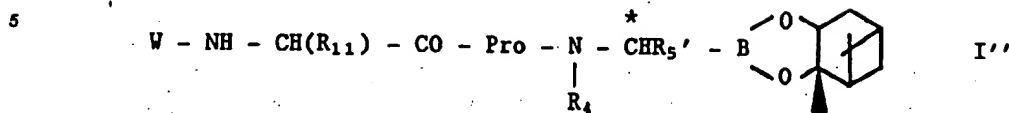
More preferred compounds are thrombin inhibitors of formula I in which Y is a sequence of two amino acids, of which the N-terminal amino acid is the unnatural amino acid and the other amino acid is L-proline (Pro), of formula



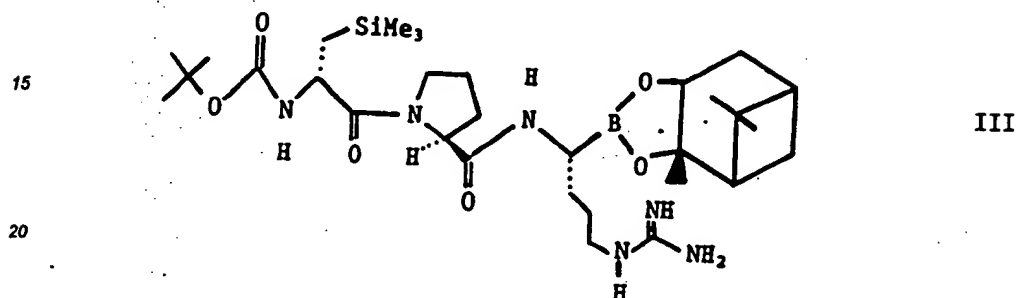
These more preferred compounds have the formula I'



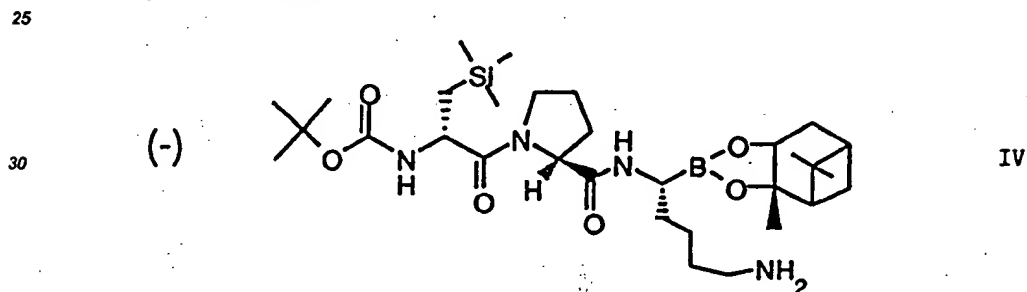
in which W, R₄, R₅, R₁₁, Q₁, and Q₂ are as defined above.
 Particularly preferred compounds are those of formula I'



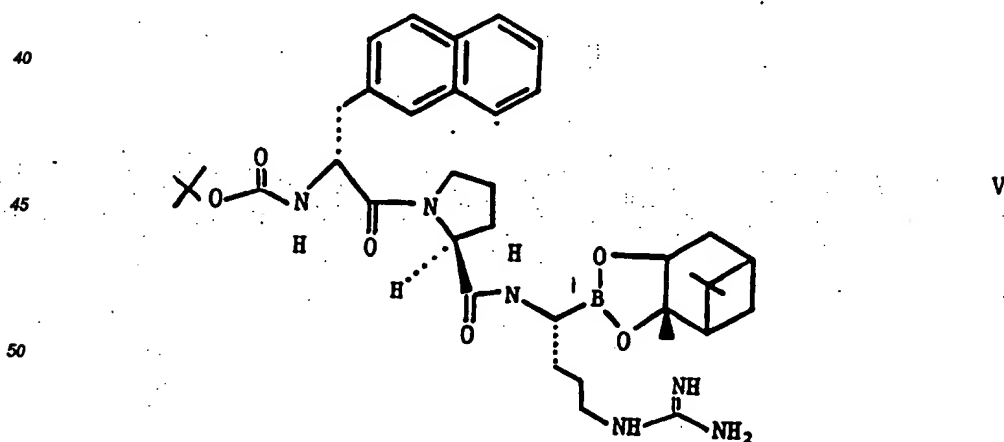
10 in which W, R₄, R_{5'} and R₁₁ are as defined above.
 The most preferred compounds are the compound of formula III



the compound of formula IV



and the compound of formula V

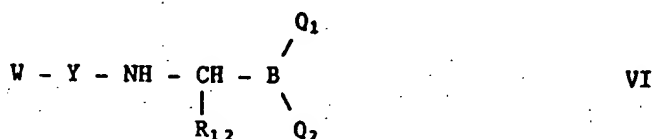


55 A peptide is considered to have an affinity for the active site of trypsin-like proteases if the particular peptide has a value of 10^{-3} M or lower for the dissociation constant.

The compounds of formula I in which X is -NH-C(NH)-NH₂ may be prepared by reacting a compound of formula I in which X is -NH₂ with cyanamide in an organic solvent under strong acid conditions. The compounds

in which X is $-NH_2$ may in turn be produced by hydrogenation of a compound of formula I in which X is $-N_3$. Hydrogenation may be carried out under standard conditions using for example a Pd/C catalyst.

The compounds of formula I in which X is $-N_3$ may be produced by reaction of a compound of formula VI

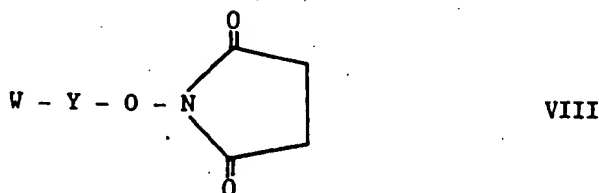


in which W, Y, Q_1 and Q_2 are defined above and R_{12} is $-A-Br$ wherein A is defined above with sodium azide in a polar aprotic solvent such as dimethyl sulfoxide. Compounds of formula I in which X is an alkylthio group may be produced by the reaction of a compound of formula VI with a thiol in the presence of an organic base such as guanidine.

The intermediates of formula VI, as well as compounds of formula I in which X is $-\text{Si}(\text{CH}_3)_3$ or an alkoxy group may be obtained by the reaction of a compound of formula VII



in which Q_1 and Q_2 are defined above and R_{13} is $-A-Br$, $A\text{-Oalkyl}$ or $-A\text{-Si}(\text{CH}_3)_3$ wherein A is defined above with $\text{LiN}[\text{Si}(\text{CH}_3)_3]_2$, followed by hydrolysis with 3 mole equivalents of acid and coupling with a protected peptide of formula VIII



wherein W and Y are defined above

The reaction is preferably carried out in a dry aprotic polar solvent, for example tetrahydrofuran, at a temperature between -78°C and room temperature.

The intermediates of formula VII may be obtained by the method of Matteson *et al.*, *Organometallics* **3** 1284-8 (1984).

The protected peptide of formula VIII may be prepared by methods which are conventional in peptide chemistry, starting from the desired unnatural amino acid. Such amino acids either are commercially available (for example the amino acid in which R_{11} is the group c) or may be prepared by methods analogous to those described in the literature, e.g. *Angew. Chem.* **93**, 793 (1981) and *J. Am. Chem. Soc.* **109**, 6881 (1987).

The compounds of formula I are useful as inhibitors of trypsin-like proteases and may be used in vitro for diagnostic and mechanistic studies of these enzymes. Furthermore, because of their inhibitory action they are indicated for use in the prevention or treatment of diseases caused by an excess of an enzyme in a regulatory system, for example control of the coagulation of fibrinolysis system.

Those compounds of the invention which are thrombin or factor Xa inhibitors have anti-thrombogenic properties and may be employed when an anti-thrombogenic agent is needed. Generally, these compounds may be administered orally or parenterally to a host to obtain an anti-thrombogenic effect. In the case of larger mammals such as humans, the compounds may be administered alone or in combination with pharmaceutical carriers or diluents at a dose of from 0.02 to 15 mg/Kg of body weight and preferably 1-10 mg/Kg to obtain the anti-thrombogenic effect, and may be given as single doses or in divided doses or as a sustained release formulation. When an extracorporeal blood loop is to be established for a patient, 0.1-1 mg/Kg may be administered intravenously. For use with whole blood from 1-10 mg per liter may be provided to prevent coagulation. Pharmaceutical diluents are well known and include sugars, starches and water which may be used to make

tablets, capsules, injectable solutions and the like. The compounds of the invention may be added to blood for the purpose of preventing coagulation of the blood in blood collecting or distribution containers, tubing or implantable apparatus which comes in contact with blood.

The advantages of the compounds of the invention include oral activity, rapid onset of activity and low toxicity. In addition, these compounds may have special utility in the treatment of individuals who are hypersensitive to compounds such as heparin.

In the following examples, the symbols have the following meanings:

Z	= benzyloxycarbonyl
Boc	= t-butyloxycarbonyl
10 Ac	= acetyl
MeOH	= methyl alcohol
EtOAc	= ethyl acetate
DCC	= dicyclohexylcarbodiimide
HONSu	= N-hydroxy-succinimide
15 OPin	= pinanediol
THF	= tetrahydrofuran
n-Bu	= n-butyl
Np	= p-nitrophenyl
TLC	= thin layer chromatography
20 Bzl	= benzyl
Baa	= -NH-CH-(CH ₂ CH ₂ CH ₂ Br)-
TMSal	= trimethylsilylalanine
Adal	= adamantylalanine
Naphal	= 2-naphthylalanine
25 BoroOrn	= -NH-CH-(CH ₂ CH ₂ CH ₂ NH ₂)B-
BoroArg	= -NH-CH-[CH ₂ CH ₂ CH ₂ NHC(NH)NH ₂]B-
Adgly	= 1-adamantylglycine
BoroPro-	= analog of proline in which the -COOH
OPin	= group is replaced by B-Opin
30 BoroLys	= -NH-CH-(CH ₂ CH ₂ CH ₂ CH ₂ NH ₂)B-
BoroHArg	= -NH-CH-(CH ₂ CH ₂ CH ₂ CH ₂ NHC(NH)NH ₂)B-
BoroMpg	= -NH-CH-(CH ₂ CH ₂ CH ₂ OCH ₃)B-
p-OH-Me-Phal	= p-hydroxymethyl-phenylalanine
p-TBDPS-O-Me-Phal	= p-tert.butyl-diphenyl-silyl-oxy- methyl-phenylalanine

35 The kinetic parameters K_i , k_{on} and k_{off} are determined by the inhibition of the enzyme catalysed hydrolysis of a peptide-arg-pNA. This hydrolysis yields p-nitroaniline, and its time-dependent release, quantified by the optical density measured at 405 nm, determines the rates of the inhibited and uninhibited reactions.

Kinetic measurements are performed on a 96-microwell plate in combination with a single-cell kinetic reader. Active-site titrated human thrombin is dissolved in 0.1 M phosphate buffer, pH 7.4, containing 0.1 M NaCl and 0.1% bovine serum albumin to a stock solution containing 400 nM active enzyme. For the chromogenic assay this solution is dissolved in the same buffer to 0.4 nM. The substrate, 2AcOH-H-D-cyclohexyl-ala-arg-pNA is dissolved in the same buffer to 0.5 mM concentration. Inhibitors are first dissolved in cremophor/ethanol 1:1 and then diluted with distilled water to 1 mM stock solution. Further dilutions are performed with the phosphate buffer described above.

45 Assays are initiated by adding 100 μ l enzyme solution to a mixture containing 100 μ l inhibitor and 50 μ l substrate solution. The release of p-nitroaniline from the hydrolysis of the peptidyl p-nitroanilide substrates is followed for 30 min to 1 h by measuring the increase in optical density at 405 nm. The collected data are used to calculate the kinetic parameters in the presence and in the absence of the inhibitor. Although other mechanisms of action are not excluded, the characterization of this class of thrombin inhibitors is restricted to the two major mechanisms observed, namely to fast, reversible and slow-, tight-binding competitive inhibition. Kinetic constants of those inhibitors displaying fast, reversible binding mechanism, namely fast binding (initial rate v_0 of control > v_0 with inhibitor) at $I_i \gg E_t$ (total inhibitor concentration/total enzyme concentration) are calculated by linear regression fitting from a $1/v$ vs. inhibitor concentration $[I]$ plots. The K_i value is calculated from the horizontal intercept $K_{i, app}$ by equation (1)

$$55 \quad K_{i, app} = K_i (1 + S/K_M) \quad (1)$$

If the rate of interaction with the enzyme is slow (v_0 not affected by the inhibitor) and tight (K_i close to or lower than E_t) so that the inhibited steady-state velocity is only slowly achieved, the progress curve of pNA formation is described by

$$P = V_0 t + \frac{(V_0 - V_s)}{k_{obs}} [1 - \exp(-k_{obs} t)] \quad (2)$$

where P is the amount of pNA formed at time 't', V_0 the initial rate, V_s the rate at steady-state, and k_{obs} an apparent global reaction rate as a function of E_0 , I_0 , K_i , K_{app} , and the observed second-order rate constant (k'_{on}) for the interaction between inhibitor and enzyme. Data from slow, tight-binding inhibition measurements are fitted to equation (2) by a nonlinear regression analysis which yields estimates of k_{obs} . Values for k'_{on} , k_{off} and K_i , K_{app} are then obtained from a plot k_{obs} vs $[I]$. The value for k_{off} is given by the vertical intercept, while the values of k'_{on} and K_i are calculated from the slope and horizontal intercepts, respectively, using equation (1).

Example 1

Boc-TMSal-Pro-NH-CH[(CH₂)₃N₃]BOPin

A. Boc-D-TMSal-OH

D-TMSal ethyl ester (21.5 g, 113.7 mMol), prepared according to the procedure given in Angew. Chem. 93, 793 (1981), is dissolved in CH₂Cl₂ and a solution of an excess of Boc₂O in CH₂Cl₂ is added. After 15 hr at room temperature, 500 ml of ice-cold 0.25N hydrochloric acid is added. The organic layer is washed with 5% NaHCO₃ and brine, then is dried over Na₂SO₄ and concentrated in vacuo.

The crude material (colourless oil) is used directly in the saponification step. It is dissolved in methanol, cooled to 0°, mixed with 510 ml of 1N NaOH and stirred at 0° for 3 hr. After acidification to pH1 with 1N HCl, the mixture is extracted several times with ether. The organic layers are combined, washed with brine, dried over Na₂SO₄ and concentrated in vacuo. The product (29.7 g oil) is used in the next step without further purification.

B. Boc-D-TMSal-Pro-ONSu

Boc-D-TMSal-OH (29.7 g, 113.7 mMol) and p-nitrophenol (19.0 g, 136.3 mMol) are dissolved in EtOAc. After cooling to 0°, DCC (23.4 g, 113.6 mMol) is added and the mixture is stirred for 1 hr at 0° and then for 15 hr at room temperature. The precipitate is filtered off and washed with EtOAc and the filtrate is concentrated in vacuo. The resulting oil is purified by flash chromatography (9:1 hexane/EtOAc) to give the desired Boc-D-TMSal-ONp as white crystals.

Boc-D-TMSal-ONp (51.6 g, 113.7 mMol) is dissolved in THF and an aqueous solution of equimolar amounts of proline and Et₃N is added. After 20 hr at room temperature, the THF is removed in vacuo and the aqueous residue is diluted with water and then extracted several times with EtOAc. The pH of the aqueous layer is adjusted to 3 by adding 10% citric acid. The resulting oily product is extracted several times with EtOAc. The combined organic layers are washed with brine, dried over Na₂SO₄ and concentrated in vacuo. The colourless oil is recrystallized from ether/hexane to give the dipeptide Boc-D-TMSal-Pro-OH as a white crystalline compound mp: 176°C.

The resulting dipeptide (26.0 g, 72.5 mMol) is dissolved in EtOAc. After cooling to 0°, HONSu (9.8 g, 85.5 mMol) and DCC (14.9 g, 72.3 mMol) are added. The mixture is stirred for 3 hr at 0° and then for an additional 15 hour at room temperature. The mixture is recooled to 0°, the dicyclohexylurea is filtered off and washed several times with EtOAc. The filtrate is washed with aqueous 0.1 M Na₂CO₃ and then with aqueous 2% KHSO₄.

After drying over Na₂SO₄ and concentration in vacuo, Boc-D-TMSal-Pro-ONSu is obtained as a white foam.

C) Boc-D-TMSal-Pro-Baa-OPin

This procedure is a one-pot 3-step procedure which comprises the *in situ* formation of a chiral α-(bis-trimethylsilyl)amido boronate, the hydrolysis of the two trimethylsilyl groups and the coupling of the so formed α-amino boronate with the active ester (Boc-D-TMSal-Pro-ONSu) prepared in step B. The entire sequence of reactions is carried out under an argon atmosphere. The chiral α-chloro boronate ((+)-Pinanediol-(S)-1-chloro-4-bromo-butane-1-boronate) (1.75 g, 5.0 mMol) is dissolved in 2.5 ml THF and is added to a precooled (-78°) solution of lithium hexamethyldisilazane (5 ml of a 1.0 M solution in hexane, 5.0 mMol). After stirring for 30 minutes at -78°, the solution is warmed up overnight to room temperature. After recooling to -78°, 3 mol equivalents of HCl in dioxane are added. The mixture is warmed up to room temperature and is stirred for 2 hr at this temperature. After recooling to -20°, a solution of the active ester of step B (2.28 g, 5.0 mMol) in 6 ml CH₂Cl₂ is added, followed by the addition of 1.39 ml (10.0 mMol) of triethylamine.

The mixture is stirred for 1 hr at -13°, warmed up to room temperature and stirred for 2 hr at this temperature. The mixture is filtered, the filtrate is concentrated in vacuo, the residue is diluted with ether and washed with 2N HCl, 5% NaHCO₃ and brine. The organic layer is dried over Na₂SO₄ and concentrated in vacuo. The residue crystallizes on standing to give the desired chiral product boronate as a white crystalline compound.

5 D) Boc-D-TMSal-Pro-NH-CH((CH₂)₃N₃)BOPin

Boc-D-TMSal-Pro-Baa-OPin, the product of step C (804 mg, 1.2 mMol) is dissolved in 13 ml of DMSO and sodium azide (156 mg, 2.4 mMol) is added. The mixture is stirred for 3 hours at room temperature. Ether / ice
10 water is added, and immediately white crystals are precipitated out of the mixture. The white precipitate is filtered off and washed with ether to give 0.6 g of the azide as a white crystalline compound.

Example 2

15 Boc-D-TMSal-Pro-BoroOrn-OPin

The azide of Example 1 (569 mg, 0.9 mMol) is dissolved in 25 ml EtOAc and is hydrogenated in the presence of 0.5 g of 10% Pd / C. After 2.5 hr, catalyst is removed and the solution is concentrated in vacuo to yield a white foam, which is recrystallized from EtOAc / ether to give the desired product as a white crystalline
20 compound, m.p.: 200 - 202°, [α]_D²⁰ = - 11.6° (c = 0.5 in MeOH).

Example 3

Boc-D-TMSal-Pro-boroArg-OPin (benzene sulfonate)

25 Boc-D-TMSal-Pro-boroOrn-OPin of Example 2 (250 mg, 0.412 mMol) is dissolved in 2 ml ethanol. Benzene sulphonic acid (65.2 mg, 0.412 mMol) is added. After stirring for 15 minutes at room temperature, cyanamide (86.6 mg, 2.06 mMol) is added and the mixture is heated under reflux. The progress of the reaction is monitored by RP-TLC in which the disappearance of the ninhydrin spot for the amine starting material and the appearance
30 of the Sakaguchi streak of the product is observed. After 7 days, amine can no longer be detected and the solution is concentrated in vacuo. The residue is dissolved in MeOH and is chromatographed on a 5 x 55 cm column of Sephadex LH-20 with MeOH.

The desired product is obtained as a white foam,
[α]_D²⁰ = - 45.3° (c = 1 in CH₂Cl₂).

35 Example 4

Boc-D-TMSal-Pro-NH-CH((CH₂)₃N₃)B-OPin

40 A) (+)-pinanediol-(S)-1-chloro-5-bromo-pentane-1-boronate

4-Bromo-1-butene (20.8 ml, 203.3 mMol) is reacted with catecholborane (24.4 g, 203.3 mMol) at 100°C over 16 hr. The crude product is distilled in vacuo to give 4-bromo-butane-1-boronate as a white crystalline compound.

45 (+)-Pinanediol (27.7 g, 163 mMol) is dissolved in THF and the above synthesized 4-bromo-butane-1-boronate (41.6 g, 163 mMol) is added. After 1 hr at room temperature, the THF is removed in vacuo and the residue is purified by flash chromatography (90:10 hexane/EtOAc) to give (+)-pinanediol-4-bromo-butane-1-boronate as a colourless oil.

The desired (+)-pinanediol-(S)-1-chloro-5-bromo-pentane-1-boronate is prepared according to the procedure given in Organometallics 3, 1284 (1984). Therefore methylenechloride (9.8 ml) in THF is cooled to -100°C and n-butyllithium (71.6 ml 1.6 M solution, 114.5 mMol) is added over 20 min. After 15 min at -100°C, a cold (-78°C) solution of (+)-pinanediol-5-bromo-pentane-1-boronate (32.8 g, 104.1 mMol) in THF is added. After additional 1 hr at -100°C anhydrous ZnCl₂ (7.1 g, 52.0 mMol) in THF is added. After additional 15 min at -100°C the reaction mixture is warmed to room temperature and stirred for 2 hr at this temperature. The solvent
55 is removed in vacuo, the residue is diluted with hexane/water and is extracted several times with hexane. After drying over Na₂SO₄ and removal of the solvent in vacuo, (+)-pinanediol-(S)-1-chloro-5-bromo-pentane-1-boronate is obtained as a yellow oil which is used directly in the next step without further purification.

B) Boc-D-TMSal-Pro-NH-CH((CH₂)₄Br)B-OPin

A solution of LiN(SiMe₃)₂ (65.2 ml 1.0 M solution, 65.2 mMol) in THF is cooled to -78°C. The α-chloro-boronate of step A) (23.7 g, 65.2 mMol) in THF is added. After stirring for 1 hr at -78°C, the mixture is stirred for 15 hr at room temperature.

After this periode, the reaction mixture is recooled to -78°C, HCl (29.8 ml 6.56 N solution, 196 mMol) in dioxan is added, the solution is stirred for 45 min at -78°C and then for 2 hr at room temperature. The mixture is cooled to -15°C, Boc-TMSal-Pro-ONSu (29.7 g, 65.2 mMol) of Example 1 in CH₂Cl₂ is added before triethylamine (18.1 ml, 130.4 mMol) is added to start the coupling reaction. After stirring at -15°C for 1 hr, the mixture is stirred for 2 hr at room temperature. The mixture is filtered over Hyflo and is concentrated in vacuo. The residue is diluted with ether/water and is extracted several times with ether. After drying over Na₂SO₄ and concentration in vacuo the desired Boc-D-TMSal-Pro-NH-CH((CH₂)₄Br)B-OPin is obtained after crystallization from ether/hexane as a white crystalline compound, mp: 74°C.

C) Boc-D-TMSal-Pro-NH-CH((CH₂)₄N₃)B-OPin

The product of step B) (33.3 g, 48.6 mMol) is dissolved in DMSO and sodium azide (6.3 g, 97.2 mMol) is added. The mixture is stirred for 6 hr at room temperature. Ether/ice water is added, and the mixture is extracted several times with ether. After drying over Na₂SO₄ and concentration in vacuo the resulting oil is crystallized to give Boc-D-TMSal-Pro-NH-CH((CH₂)₄N₃)B-Pin as a white crystalline compound, mp: 69-70°C; α_D = -74.4° (c=1.0 in MeOH).

Example 5Boc-D-TMSal-Pro-BoroLys-OPin

The azide of Example 4 (22.0 g, 34.0 mMol) is dissolved in EtOAc and is hydrogenated in the presence of 4.0 g of 10% Pd/C. After 9 hr, the catalyst is removed and the solution is concentrated in vacuo. The resulting foam is dissolved in EtOAc and is crystallized to give the desired Boc-D-TMSal-Pro-BoroLys-OPin as white crystals, mp 128-129°C; α_D = -59.6° (c=1.0 in MeOH).

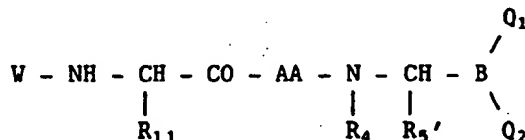
Example 6Boc-D-TMSal-Pro-BoroHArg-OPin (benzene sulfonate)

The benzene sulfonate of Boc-D-TMSal-Pro-BoroLys-OPin of Example 5 (800 mg, 1.03 mMol) is dissolved in ethanol. Cynamide (210 mg, 5.0 mMol) is added and the mixture is heated under reflux. The progress of the reaction is monitored by RP-TLC in which the disappearance of the ninhydrin spot for the amine starting material and the appearance of the Sakaguchi streak of the product is observed. After 7 days, amine can no longer be detected and the solution is concentrated in vacuo. The residue is dissolved in MeOH and is chromatographed on a 5 x 55 cm column Sephadex LH-20 with MeOH.

The desired product is obtained as a white foam, α_D = -40.8° (c=0.5 in CH₂Cl₂).

Examples 7-29

By methods analogous to those in Examples 1-6, the compounds of formula



in which R₄, R₅', R₁₁, Q₁ + Q₂ and AA have the significances shown in the following table I can be obtained

TABLE I

	Ex.	W	R ₁₁	R ₄	R ₅ '	Q ₁ +Q ₂	AA	[α] _D ²⁰	C	solvent
5	7	Boc	c)	H	-(CH ₂) ₃ -NH ₂	a)	Pro	-27,8°	0,5	EtOH
	8	Boc	c)	H	-(CH ₂) ₃ NHC(NH)NH ₂	a)	Pro	-75,0°	1,0	CH ₂ Cl ₂
	9	Boc	e)	H	-(CH ₂) ₄ N ₃	a)	Pro	-64,7°	0,51	CH ₂ Cl ₂
10	10	Boc	e)	H	-(CH ₂) ₂ SiMe ₃	a)	Pro	-57,0°	0,5	CH ₂ Cl ₂
	11	Boc	d)	H	-(CH ₂) ₂ SiMe ₃	a)	Pro	-59,3°	1,0	CH ₂ Cl ₂
	12	Ac	c)	H	-(CH ₂) ₃ NHC(NH)NH ₂	a)	Pro	-80,2°	0,5	CH ₂ Cl ₂
15	13	Boc	e)	H	-(CH ₂) ₃ NHC(NH)NH ₂	a)	Pro	-42,3°	0,5	CH ₂ Cl ₂
	14	Boc	c)	H	-(CH ₂) ₄ NHC(NH)NH ₂	a)	Pro	-18,8°	0,5	EtOH
	15	Boc	e)	H	-(CH ₂) ₄ NHC(NH)NH ₂	a)	Pro	-61,7°	0,41	CH ₂ Cl ₂
	16	Boc	e)	H	-(CH ₂) ₄ NH ₂	a)	Pro	-52,4°	0,54	CH ₂ Cl ₂
20	17	Boc	d)	H	-(CH ₂) ₃ NHC(NH)NH ₂	b)	Pro	-35,8°	1,0	EtOAc
	18	Boc	e)	H	-(CH ₂) ₃ -NH ₂	a)	Pro	-10,6°	0,5	CH ₂ Cl ₂
	19	Boc	d)	H	-(CH ₂) ₃ NHC(NH)NH ₂	b)	Gly	+ 8,8°	0,5	EtOAc
25	20	Ac	c)	H	-(CH ₂) ₃ -NH ₂	a)	Pro	-78,6°	0,75	CH ₂ Cl ₂
	21	Boc	e)	H	-(CH ₂) ₃ -NH ₂	b)	Gly	-4,4°	0,5	CH ₂ Cl ₂
	22	Boc	f)	H	-(CH ₂) ₃ -N ₃	a)	Pro	-76,0°	0,5	CH ₂ Cl ₂
	23	Boc	f)	H	-(CH ₂) ₃ -NH ₂	a)	Pro	-25,4°	0,5	CH ₂ Cl ₂
30	24	Boc	f)	H	-(CH ₂) ₃ NHC(NH)NH ₂	a)	Pro	-34,8°	0,5	CH ₂ Cl ₂
	25	Boc	d)	H	-(CH ₂) ₄ NHC(NH)NH ₂	a)	Gly	-14,7°	1,0	MeOH
	26	Boc	d)	H	-(CH ₂) ₄ NH ₂	a)	Gly	-30,6	0,5	EtOH
35	27	Boc	d)	H	-(CH ₂) ₄ NH ₂	a)	Asp	-18,4°	0,5	MeOH
	28	H	d)	H	-(CH ₂) ₃ NH ₂	a)	Pro	-53,6	0,32	MeOH
	29	Boc	d)	H	-(CH ₂) ₄ OC ₂ H ₅	a)	Pro	-54,2	0,5	MeOH

40 **Example 30****Boc-D-TMSal-Pro-BoroMpg-OPin****A) (+)-Pinanediol-(S)-1-chloro-4-methoxy-butane-1-boronate**

45

3-Methoxy-1-propene (6.0 g, 83.3 mMol) is reacted with catecholborane (10.0 g, 83.3 mMol) at 100°C over 24 hr. The crude product is distilled in vacuo to give 3-methoxy-propane-1-boronate as a colourless oil.

(+)-Pinanediol (10.6 g, 62.5 mMol) is dissolved in THF and the above synthesized 3-methoxy-propane-1-boronate (12.0 g, 62.5 mMol) is added. After 1 hr at room temperature, the THF is removed in vacuo and the residue is purified by flash chromatography (80:20 hexane/EtOAc) to give (+)-pinanediol-3-methoxy-propane-1-boronate as a colourless oil.

The desired (+)-pinanediol-(S)-1-chloro-4-methoxy-butane-1-boronate is prepared according to the procedure given in Organometallics 3, 1284 (1984). Therefore methylenechloride (2.2 ml) in THF is cooled to -100°C and *n*-butyllithium (13.8 ml 1.6 M solution, 22.0 mMol) is added over 20 min. After 15 min at -100°C, a solution of (+)-pinanediol-3-methoxy-propane-1-boronate (5.04 g, 20 mMol) in THF is added followed by anhydrous ZnCl₂ (1.42 g, 10.0 mMol). After additional 15 min at -100°C the reaction mixture is warmed to room temperature and stirred for 2 hr at this temperature. The solvent is removed in vacuo, the residue is diluted with ether and washed with water. The organic layer is dried over Na₂SO₄ and is concentrated to give an oil

which is purified by flash chromatography (9:1 hexane/EtOAc) to give the desired (+)-pinanediol-(S)-1-chloro-4-methoxybutane-1-boronate as a colourless oil.

B) Boc-D-TMSal-Pro-BoroMpg-OPin

5

A solution of $\text{LiN}(\text{SiMe}_3)_2$ (5 ml 1.0 M solution, 5.0 mMol) in THF is cooled to -78°C . The α -chloro-boronate of step A) (1.53 g, 5.0 mMol) in THF is added. After stirring for 1 hr at -78°C , the mixture is stirred for 15 hr at room temperature. After this period, the reaction mixture is recooled to -78°C , HCl (2.7 ml 5.65 N solution, 15.0 mMol) in dioxane is added, the solution is stirred for 30 min at -78°C and then for 2 hr at room temperature. The mixture is cooled to -15°C , Boc-TMSal-Pro-ONSu (2.28 g 5.0 mMol) of Example 1 in CH_2Cl_2 is added before triethylamine (1.39 ml, 10.0 mMol) is added to start the coupling reaction. After stirring at -15°C for 1 hr, the mixture is stirred for 2 hr at room temperature. The mixture is filtered over Hyflo and is concentrated in vacuo. The residue is diluted with EtOAc and is washed with 0.2 N HCl, 5% NaHCO_3 and finally with brine. After removal of the solvent, an oil is obtained which is purified by flash chromatography (EtOAc) to give Boc-D-TMSal-Pro-BoroMpg-OPin as a white foam, $\alpha_D = -48.8^\circ$ ($c=0.25$ in CH_2Cl_2).

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Example 31

Boc-D-(p-(TBDPS-O)methyl)Phal-Pro-BoroOm-OPin

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A) Boc-D-(p-((1,1-dimethylethyl)diphenyl-silyl)oxy)methylphenylalanine

In order to selectively reduce the azido group in the substrate, thiophenol (7.27 g, 66.0 mMol) is added to a suspension of SnCl_2 (3.12 g, 16.5 mMol) in CH_2Cl_2 . Triethylamine (6.8 ml, 49.5 mMol) is added and a yellow solution is obtained. Boc-anhydride (4.8 g, 22.0 mMol) is added before (3(2S), 4S-3-(2-azido-3-(p-((1,1-dimethyl)di-phenyl-silyl)oxy-methyl)phenyl-1-oxo-propyl)-4-(phenyl-methyl)-2-oxazolidinone (%de>95; 6.8 g, 11.0 mMol), prepared according to the procedure given in J. Am. Chem. Soc. 109 6881 (1987), is added as a solution in CH_2Cl_2 . After stirring for 2.5 hr at room temperature, the mixture is diluted with EtOAc/2 N NaOH and filtered over Hyflo. The organic layer is washed with 2% aqueous NaHSO_4 , 5% aqueous NaHCO_3 and finally with brine. After drying over Na_2SO_4 and concentration in vacuo the obtained yellow oil is purified by flash chromatography to give (3(2S),4S)-3-(2-(((tert.-butoxy)-carbonyl)amino)-3-(p-(((1,1-dimethylethyl)diphenyl-silyl)oxy)-methyl)-phenyl-1-oxopropyl)-4-(phenylmethyl)-2-oxazolidinone as a white foam. This compound (2.0 g, 2.88 mMol) is dissolved in THF/water and is hydrolyzed with in situ generated LiOOH (5.76 mMol) at 0°C . After 1.75 hr at 0°C , Na_2SO_3 (1.25 g, 9.9 mMol) in water is added. THF is removed in vacuo, the pH of the residue is adjusted to 1-2 and the mixture is extracted 3 times with EtOAc. The combined organic layers are washed with water, dried over Na_2SO_4 and are concentrated in vacuo. After crystallization from hexane/ether the oxazolidinone is obtained as white crystals. The filtrate is concentrated in vacuo to give the desired title compound as a white foam.

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B) Boc-D-(p-(((1,1-dimethylethyl)diphenyl-silyl)oxy)-methyl)-phenylalanine-Pro-ONSu

DCC (0.59 g, 2.88 mMol) is added to a mixture of the title compound of step A) (1.6 g, 2.88 mMol) and p-nitrophenol (0.43 g, 3.12 mMol) in EtOAc at 0°C . The reaction mixture is stirred for 16 hr at room temperature. After cooling to 0°C , the precipitate is filtered off and washed with cold EtOAc. The filtrate is concentrated in vacuo. The resulting oil (Boc-D-(p-TBDPS-O-Me)-Phal-ONp) is used in the next step without further purification. Boc-D-(p-TBDPS-O-Me)-Phal-ONp (2.2 g, 2.88 mMol) is dissolved in THF and an aqueous solution of L-proline (365 mg, 3.17 mMol) and Et_3N (0.88 ml, 6.33 mMol) is added. After 15 hr at room temperature the THF is removed in vacuo. The pH is adjusted to 3 by adding 10% citric acid. The resulting oily product is extracted several times with EtOAc. The combined organic layers are washed with brine, dried over Na_2SO_4 and concentrated in vacuo. The colourless oil is purified by flash chromatography (9:1 $\text{CH}_2\text{Cl}_2/\text{EtOH}$ to eluate p-nitrophenol, then 80:20 $\text{CH}_2\text{Cl}_2/\text{EtOH}$) to give Boc-D-(p-TBDPS-O-Me)-Phal-Pro-OH as a white foam. This dipeptide (1.3 g, 2.06 mMol) is dissolved in EtOAc. After cooling to 0°C , HONSu (220 mg, 2.47 mMol) and DCC (330 mg, 2.06 mMol) are added. The mixture is recooled to 0°C , the dicyclohexylurea is filtered off and washed several times with cold EtOAc. The filtrate is washed with aqueous 0.1 M Na_2CO_3 , 8% NaHSO_4 and then with brine. After drying over Na_2SO_4 and concentration in vacuo the title compound Boc-D-(p-TBDPS-O-Me)-Phal-Pro-ONSu is obtained as a white foam.

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C) Boc-D-(p-TBDPS-O-Me)-Phal-Pro-Baa-OPin

The title compound is obtained by using the analogous one-pot 3-step procedure described for the synthesis of Boc-D-TMSal-Pro-Baa-OPin in Example 1/C. Therefore the intermediate α -amino-boronate, which results from the reaction of the chiral α -chloro-boronate ((+)-pinanediol-(S)-1-chloro-4-bromo-butane-1-boronate) (659 mg, 2.0 mMol) with $\text{LiN}(\text{SiMe}_3)_2$ (2.0 mMol) and hydrolysis with HCl, is reacted with the active ester of step B) (1.45 g, 2.0 mMol) in the presence of Et_3N (4.0 mMol) to give the title compound which is purified by flash chromatography (1:1 hexane/EtOAc).

D) Boc-D-(p-TBDPS-O-Me)-Phal-Pro-BoroOrn-OPin

The product of step C) (680 mg, 0.72 mMol) is dissolved in DMSO and sodium azide (94 mg, 1.44 mMol) is added. The mixture is stirred for 4 hr at room temperature. Ether/water is added, and immediately white crystals are precipitated out of the mixture. The white precipitate is filtered off and washed with water to give Boc-D-(p-TBDPS-O-Me)-Phal-Pro-NH-CH((CH₂)₃N₃)B-OPin as a white crystalline compound. This azide (272 mg, 0.3 mMol) is dissolved in EtOAc and is hydrogenated in the presence of Lindlar-Catalyst. After 8 hr, the catalyst is removed and the solution is concentrated in vacuo. The crude product is purified by flash chromatography (EtOAc then EtOH) to give the desired title compound as a white foam, $\alpha_D = -32.4^\circ$ (c=0.25 in MeOH).

Example 32

Boc-D-(p-OH-Me)-Phal-Pro-BoroOrn-OPin

The boro-ornithin of Example 31 (132 mg, 0.15 mMol) is dissolved in THF and is reacted with $n\text{-Bu}_4\text{NF}$ (0.3 ml 1.1 M solution, 0.3 mMol). After 45 min at room temperature ice water is added and the resulting mixture is extracted several times with EtOAc. The combined organic layers are dried over Na_2SO_4 and concentrated in vacuo. The resulting oil is purified by a short chromatography (EtOAc then EtOH) to give the desired title compound as a white foam, $\alpha_D = -34.0^\circ\text{C}$ (c=0.1 in MeOH).

Example 33

Boc-D-TMSal-Adgly-boroPro-OPin

A. L-1-Adamantylglycine

(3(2S),4S)-3-(2-Azido-2-adamant-1-yl-1-oxoethyl)-4-(phenylmethyl)-2-oxazolidone (% de > 95; 9.86 g, 25.0 mMol), prepared according to the procedure given in J. Am. Chem. Soc. 109, 6881 (1987), is dissolved in 320 ml of a mixture of THF/H₂O (3:1), cooled to 0°C, mixed with 4 equiv. of hydrogen peroxide and 2.0 equiv. of LiOH. The resulting mixture is stirred at 0°C until the substrate has been consumed (30 min.), and the peroxide (percarboxylate) is quenched at 0°C with 10% excess of 1.5 N aq Na_2SO_3 . After buffering with aqueous NaHCO_3 (pH 9-10) the mixture is extracted several times with EtOAc to remove the oxazolidinone chiral auxiliary. The product carboxylic acid is isolated by EtOAc extraction of the acidified (pH 1-2) aqueous phase, drying over Na_2SO_4 and concentration in vacuo. The desired (S)-azido-adamant-1-yl-acetic acid is isolated as white crystals (5.29 g) and is used in the next step without further purification. 2(S)-Azido-adamant-1-ylacetic acid (5.29 g, 22.5 mMol) is dissolved in 110 ml of EtOH and 11.3 ml of 2 N HCl, and is hydrogenated in the presence of 0.7 g of 10% Pd/C. After 2.5 h, catalyst is removed and the solution is concentrated in vacuo to yield the desired amino acid as the hydrochloride salt. The obtained hydrochloride is suspended in 40 ml of H₂O and is treated with 1.9 g of solid NaHCO_3 . The obtained amino acid is filtered off and washed several times with water. After drying in vacuo, L-1-adamantyl-glycine is obtained as a white crystalline compound, $[\alpha]_D^{20} = +3.0$ (c = 1.0 in MeOH).

B. Boc-D-TMSal-Adgly-ONSu

Boc-D-TMSal-ONp (7.71 g, 20.2 mMol) of example 1 is dissolved in THF and an aqueous solution of equimolar amounts of 1-adamantyl-glycine and Et_3N is added. After 20 h at room temperature the THF is removed in vacuo and the aqueous residue is diluted with 150 ml of 0.1 N HCl, and then extracted several times with EtOAc. The combined organic layers are washed with brine, dried over Na_2SO_4 and concentrated in vacuo.

The oily product is chromatographed on silica gel (CH_2Cl_2) to give the dipeptide Boc-D-TMSal-Adgly-OH as an oil. Boc-D-TMSal-Adgly-OH (6,9 g, 15,2 mMol) is dissolved in 80 ml of EtOAc. After cooling to 0°C , HONSu (2,1 g, 18,0 mMol) and DCC (3,1 g, 15,2 mMol) are added. The mixture is stirred for 3h at 0°C and then for an additional 15h at room temperature. The mixture is recooled to 0°C , the dicyclohexylurea is filtered off and washed with EtOAc. The filtrate is washed with aqueous 0,1 M NaHCO_3 and then with aqueous 2% KHSO_4 . After drying over Na_2SO_4 and concentration in vacuo, Boc-D-TMSal-Adgly-ONSu (7,2 g) is obtained as a white foam.

C. Boc-D-TMSal-Adgly-boroPro-OPin

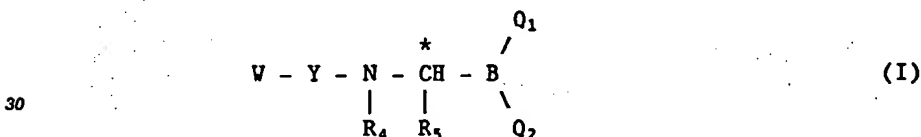
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The title compound is obtained by using the analogous one-pot 3-step procedure described for the synthesis of Boc-D-TMSal-Pro-Baa-OPin in example 1/C. Due to the low reactivity of the sterically hindered active ester of step B (2,7 g, 5,0 mMol), the intermediate α -amino boronate, which results from the reaction of the chiral α -chloro-boronate ((+)-Pinanediol-(S)-1-chloro-4-bromo-butane-1-boronate) (1,7 g, 5,0 mMol) with lithium hexamethyl-disilazane (5,0 mMol) and hydrolysis with HCl, cyclizes to the boroproline derivative, which reacts then with the active ester of step B to give the unexpected Boc-D-TMSal-Adgly-boroPro-OPin as the major product. Flash chromatography (2:1 hexane/EtOAc) of the crude product yields the title compound (0,48g) as a white foam, which is further purified by recrystallization from ether/hexane to give the desired product Boc-D-TMSal-Adgly-boroPro-OPin as a white crystalline compound.

mp: $187-188^\circ\text{C}$, $[\alpha]_D^{20} = +2,8^\circ$ ($c = 1,0$ in CH_2Cl_2).

Claims

1. A compound of formula I



wherein:

W is hydrogen or an N-protecting group

Y is a sequence of n amino acids such that the n+1 amino acid peptide Y-Lys or Y-Arg has an affinity for the active site of a trypsin-like protease; where n is an integer of from 1 to 10 and in which at least one amino acid is an unnatural amino acid having a hydrophobic side chain;

Q_1 and Q_2 are the same or different and are selected from -OH, -COR₁, -CONR₁R₂, -NR₁R₂ or -OR₃ or Q_1 and Q_2 taken together form a diol residue;

R_1 , R_2 and R_3 which may be the same or different, are C_{1-10} alkyl, C_{6-10} aryl, C_{6-10} aralkyl or phenyl substituted by up to three groups selected from C_{1-4} alkyl, halogen and C_{1-4} alkoxy;

R_4 is hydrogen or C_{1-10} alkyl

R_5 is a group -A-X; wherein

A is $-(\text{CH}_2)_z-$ in which z is 2,3,4 or 5;

$-\text{CH}(\text{CH}_3)-(\text{CH}_2)_2-$; $-\text{CH}_2-\text{CH}(\text{CH}_3)-\text{CH}_2-$; $-(\text{CH}_2)_2-\text{CH}(\text{CH}_3)-$;

$-(\text{CH}_2)_2-\text{C}(\text{CH}_3)_2-$; $-\text{CH}(\text{CH}_3)-(\text{CH}_2)_3-$; $-\text{CH}_2-\text{CH}(\text{CH}_3)-(\text{CH}_2)_2-$;

$-\text{CH}_2-\text{CH}_2-\text{CH}(\text{CH}_3)-\text{CH}_2-$; $(\text{CH}_2)_3-\text{CH}(\text{CH}_3)-$; $-(\text{CH}_2)_3-\text{C}(\text{CH}_3)_2$; C_{6-10} aryl C_{6-10} aralkyl and

X is $-\text{NH}_2$, $-\text{NH}-\text{C}(\text{NH})-\text{NH}_2$, $-\text{S}-\text{C}(\text{NH})-\text{NH}_2$, $-\text{N}_3$, C_{1-4} alkoxy, C_{1-4} alkylthio or $-\text{Si}(\text{CH}_3)_3$ or

R_4 and R_5 together form a trimethylene group and the asymmetric carbon atom marked * may have the D- or L- configuration or represent any mixture of these.

2. A compound according to claim 1 in which W is $\text{H}(\text{CH}_2\text{CH}_2\text{O})_p-$, $\text{R}_6\text{CO}-$, $\text{R}_7\text{OCO}-$ or R_8SO_2- , wherein:

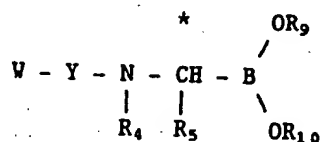
p = 3-30

$\text{R}_6 = \text{C}_{1-6}$ alkyl

$\text{R}_7 = \text{C}_{1-6}$ alkyl, phenyl, benzyl or naphthyl; and

$\text{R}_8 = \text{phenyl, naphthyl or } \text{C}_{1-4}\text{alkylphenyl}$

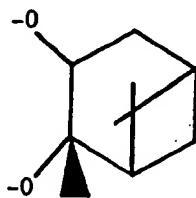
- 3. A compound according to claim 1 or 2 which is of formula Ia**



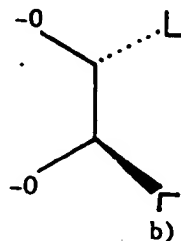
Ia

wherein: W, Y, R₄ and R₅ are as defined in claims 1 or 2 and R₉ and R₁₀ represent the residue of a dihydroxy compound.

4. A compound according to claim 1 or 2 wherein Q₁ and Q₂ together represent a group of formula (a) or (b)

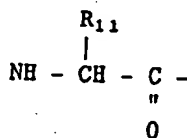


(a)



b)

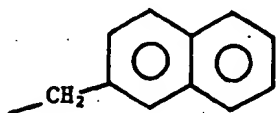
5. A compound according to any one of claims 1 to 4 wherein the unnatural amino acid is of formula II



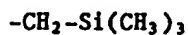
(II)

wherein R_{11} is a hydrophobic group.

- 35 6. A compound according to claim 5 wherein R₁₁ is R₁₁' and is a group of formula (c), (d), (e), (f), (g), (h) or (i)



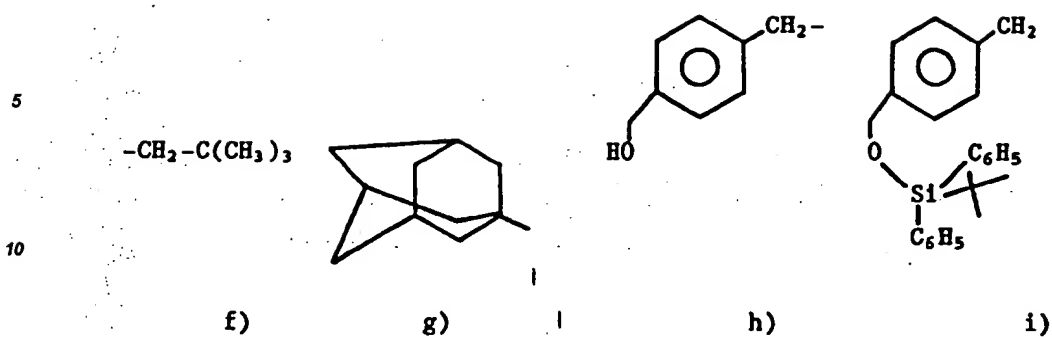
c)



d)

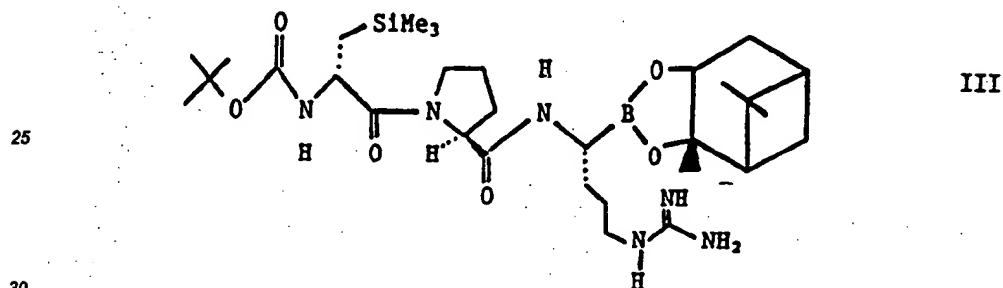


e)

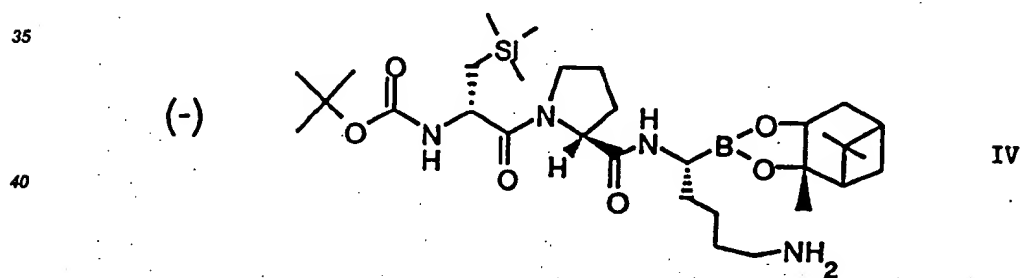


15 7. A compound according to claim 1 wherein Y is a sequence of two amino acids of which the N-terminal amino acid is the unnatural amino acid and the other amino acid is L-proline.

20 8. A compound according to claim 1 which is of formula III



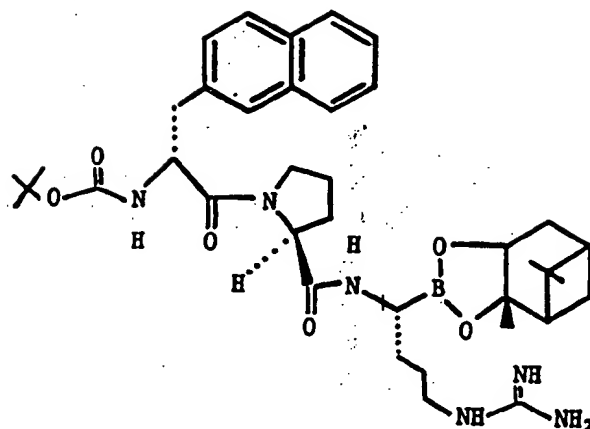
35 9. A compound according to claim 1 which is of formula IV



45 10. A compound according to claim 1 which is of formula V

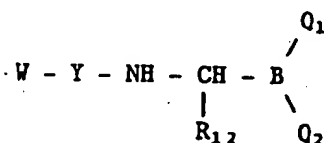
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V

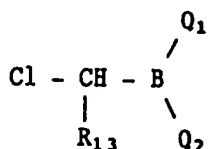
11. A process for the preparation of a compound of formula I as defined in claim 1 which comprises:
- i) when X is -NH-C(NH)-NH₂, reacting compound of formula I in which X is -NH₂ with cyanamide,
 - ii) when X is -NH₂, hydrogenating a compound of formula I in which X is -N₃;
 - iii) when X is -N₃, reacting a compound of formula VI



(VI)

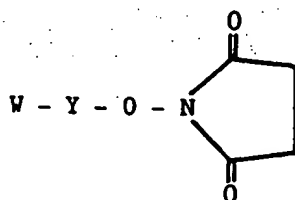
In which W, Y, Q₁ and Q₂ are defined in claim 1 and R₁₂ is -A-Br, A being as defined in claim 1, with sodium azide;

iv) when X is -Si(CH₃)₃ or O-alkyl, reacting a compound of formula VI



(VII)

In which Q₁ and Q₂ are defined in claim 1 and R₁₃ is -A-Si(CH₃)₃ or O-alkyl with LiN[Si(CH₃)₃]₂, followed by acid hydrolysis and coupling with a protected peptide of formula VIII

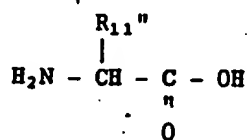


(VIII)

wherein W and Y are defined in claim 1.

- 12. Use of a compound of any one of claims 1 to 10 as inhibitors of trypsin like serine proteases.
- 13. Therapeutical composition containing a compound of any one of claims 1 to 10 together with pharmaceutically acceptable additives and/or diluents.

14. Use of a therapeutical composition according to claim 13 for inhibition of trypsin like serine proteases.
15. Use according to claims 12 and 14 characterised in that trypsin-like serine proteases are thrombin, factor Xa, kallikrein, plasma, prolyl endopeptidase and Ig A1 protease.
16. Therapeutical composition having anti-thrombogenic activity containing a compound of any one of claims 1 to 10 together with pharmaceutically acceptable additives and/or diluents.
17. Use of a compound of any one of claims 1 to 10 as inhibitors of thrombin.
18. Unnatural amino acid of formula IIa



wherein R₁₁'' is a group of formula (d), (e), (f), (g), (h) or (i) which are defined in claim 6.